Possible Role of Autoimmunity in Patients with Premature Ovarian Insufficiency

Renata Košir Pogačnik, M.D.^{1*}, Helena Meden Vrtovec, Ph.D.¹, Alenka Vizjak, Ph.D.², Alenka Uršula Levičnik, M.D.¹, Nina Slabe, M.D.¹, Alojz Ihan, Ph.D.³

- 1. Department of Obstetrics and Gynecology, University Medical Centre Ljubljana, Ljubljana, Slovenia
 - 2. Institute of Pathology, Faculty of Medicine, University of Ljubljana, Ljubljana, Slovenia
 - 3. Institute of Microbiology and Immunology, Faculty of Medicine, University of Ljubljana, Ljubljana, Slovenia

Abstract-

Background: To evaluate the involvement of immune abnormality in patients with idiopathic premature ovarian insufficiency (POI). In addition to the known etiology, autoimmune disorders may be a pathologic mechanism for POI.

Materials and Methods: Our study was a prospective controlled trial. Twenty women with POI, reasons other than autoimmune excluded, were enrolled in this study. The control group consisted of 17 healthy women. In both groups, family and personal history were taken and the levels of follicle stimulating hormone, luteinizing hormone, thyroid-stimulating hormone, prolactin, anti-Müllerian hormone, inhibin B, antithyroglobulin and antithyroid peroxidase antibodies were determined. Antiovarian antibodies and subpopulations of peripheral blood T-lymhocytes were also determined.

Results: Participants in the study group exhibited hypergonadotropichypogonadism, while high levels of follicle stimulating hormone and low levels of inhibin B and anti-Müllerian hormone were observed. In 16 (80%) patients, POI was associated in their personal and familial history with another autoimmune disease. Fifty percent of patients presented highly elevated antithyroid antibodies. The lymphocyte subset, especially B cells, was significantly higher (p=0.014), and peripheral regulatory lymphocytes CD25⁺ were significantly lower (p=0.015) in the study group than in the control group. Antiovarian antibodies were detected in 20% of patients with POI.

Conclusion: We presume that the presence of anti-ovarian antibodies together with abnormalities of cellular immunity may in some cases potentially represent the involvement of an autoimmune mechanism in idiopathic POI.

Keywords: Autoantibody, Thyroid Stimulating Antibody, Cell Immunity, Premature Ovarian Insufficiency, T-Lymphocyte

Citation: Košir Pogačnik R, Meden Vrtovec H, Vizjak A, Uršula Levičnik A, Slabe N, Ihan A. Possible role of autoimmunity in patients with premature ovarian insufficiency. Int J Fertil Steril. 2014; 7(4): 281-290.

Introduction

Premature ovarian insufficiency (POI) is characterized by hypergonadotropic amenorrhea due to cessation of ovarian function before the age of 40 years. The diagnosis is based on amenorrhea before the age of 40 associated with follicle stimulat-

ing hormone (FSH) levels >40 IU/l, detected on two occasions at least one month apart (1). POI causes female infertility, while is a significant psychosocial burden and a risk to women's health. It occurs in 1% of women, of whom 10-28% have primary and 4-18% secondary amenorrhea (2, 3).

Received: 17 Nov 2012, Accepted: 07 Apr 2013

Email: renata.kosir@inbox.com



^{*} Corresponding Address: Department of Obstetrics and Gynecology, University Medical Centre Ljubljana, Šlajmerjeva 3, SI-1000 Ljubljana, Slovenia

Although there are multiple etiologies of POI (genetic, chromosomal, infectious, and iatrogenic causes), the etiology cannot be identified in most patients and this is referred to as idiopathic POI; up to 30% of idiopathic cases may have an autoimmune cause (4). The most convincing evidence coming from the commonly observed association of POI with other autoimmune disorders (5, 6) are demonstration of anti-ovarian antibodies (AOA, 7, 8) and histological findings of ovarian tissue from affected women. Roughly, one third of POI patients have AOA and/or antithyroid antibodies in their serum (1, 9). Various organ-specific and systemic autoimmune diseases cause autoimmune ovarian insufficiency in up to 30% of women with POI (4). According to the literature, 2-10% of POI cases are known to be associated with adrenal autoimmunity (10). One of the first signs that autoimmunity may be responsible for ovarian function failure came from the observation that ovarian failure may precede the onset of Addison's disease by 8-14 years (11). Autoimmune Addison's disease seldom develops in isolation, whereas several other endocrine glands and organs are generally affected, leading to an autoimmune polyglandular syndrome (APGS, 12). Two main forms of APGS can be clinically discerned, APGS types 1 and 2. APGS type 1 is characterized by an association of mucocutaneous candidiasis, hypoparathyroidism and Addison's disease. In about 60% of cases, there is also an association with ovarian insufficiency. Blizzard et al. (13) and Irvine et al. (14) found that POI commonly presents with adrenal cytoplasmic antibodies, called steroid cell antibodies (SCA); they react with cytoplasmic antigens of other steroid-producing cells present in the ovary, testis and placenta. Alteration of lymphocytes and their specific subsets, as well as T-cell mediated injury are likely to play an important role in the pathogenesis of autoimmune oophoritis. Surface markers of peripheral blood mononuclear cells (PBMC) have been shown to be deranged in early autoimmune phases and to be persisted through the disease, even after targeted disruption (15).

The presence of pathogenic factors might accelerate the process of apoptosis and atresia of ovarian follicles during the fetal and post-natal period (16). This interpretation is based on the dogma that the number of ovarian follicles at birth is final and that there is no possibility of regeneration or renewal of reserve follicles in adulthood (17). Ex-

perimental work on animals suggests a possibility of renewal of the follicle reserve from proliferative germinal ovarian cells even after birth; verification of which is being sought in studies on human ovaries (18, 19). It has been shown that undifferentiated ovarian stem cells differentiate into structures similar to egg cells under certain laboratory conditions (20). We faced two problems: whether to accept the standard understanding of a final number of ovarian follicles or to focus on the hypothetic possibility of renewal of the follicular reserve. The aim of this study was to evaluate the involvement of immune abnormality in patients with idiopathic POI

Materials and Methods

Subjects

Our study was a prospective randomized controlled trial. The study group consisted of 20 women with POI (mean age 31.8 years, range 20-39 years) and no use of medications or oral contraceptives for at least 4 months prior to the study. The diagnosis was based on the presence of amenorrhea before the age of 40, associated with two serum FSH levels above 40 IU/l at least one month apart. All women with POI underwent karyotyping and genetic testing of the fragile X mental retardation 1 (FMR1). Women with infectious or iatrogenic causes were excluded from the study. The control group consisted of 17 healthy women volunteers. Inclusion criteria were a regular menstrual cycle, age between 18 and 39 years (mean 30.8 years), and no use of medications or oral contraceptives for at least 4 months prior to the study. They also had to be exempt from autoimmune disease or infertility problems. All women provided a complete personal and family history, with a stress on possible immune-mediated, particularly autoimmune processes (allergy, asthma, diabetes, thyroiditis, rheumatoid arthritis, andatopiceczema), and all underwent physical and vaginal ultrasound examinations.

Hormone and serological analyses

Serum levels of luteinizing hormone (LH), FSH, estradiol (E₂), prolactin (PRL), inhibin B, thyroid-stimulating hormone (TSH), anti-Müllerian hormone (AMH), antithyroglobulin (aTG) and antithyroid peroxidase antibodies (aTPO) were

measured, and immunological investigations at cellular and humoral levels were performed. The adrenocorticotropic hormone (ACTH) stimulation test was performed in the study group, only. The standard Synacthen stimulation test is clinically widely used as a sensitive screening method for symptomatic adrenal insufficiency. Each ampoule of Synacthen contains 250 µg of the active ingredient, tetracosactrin (Novartis Pharmaceuticals, North Ryde NSW, Australia). Thirty minutes after 250 µg Synacthen I.M. (Alliance Pharmaceutical Wiltshire, UK), blood cortisol was measured by electro-chemiluminescence immunoassay. A normal cortisol response to Synacthen was defined as a post-stimulation peak cortisol value of >500 nmol/l at 30 minutes.

Serum AMH in peripheral blood was determined by a Personal Lab analyser using the enzyme linked immunosorbent assay (ELISA) method with Beckman Coulter reagent. The normal range of AMH levels is 0.7-3.5 mg/l. Values below 0.3 mg/l indicate a reduced ovarian reserve. Hormones were determined on a LI-AISON analyser by quantitative direct competitive chemiluminescence immunoassays (CLIA). Each test is a modified two-step process, in which the specific antibodies of a certain hormone bind to magnetic cells. Normal ranges for the follicular phase of the cycle are as follows: FSH: 3.5-9.2 IU/l; LH: 1.1-11.6 IU/l; LH around ovulation: 17-77 IU/l; PRL: 6.2-23.5 μg/l; Ε₂: on day 3 up to 310 pmol/L (0.31 nmol/L); \tilde{E}_{3} postmenopause: 0-110 pmol/L (0-0.11 nmol/l); and TSH: 0.3-3.6 mE/l. Serum Inhibin-B cut-off level on day 3 was 45 pg/mL. Serum aTG and aTPO concentrations measured by immunoassay using direct chemiluminometric technology on an ADVIA CENTAUR analyser (Siemens Medical Solutions Diagnostics, Tarrytown, USA). The normal range for serum aTG and aTPO is <60 KE/l. Detection of AOA was by indirect immunofluorescence (IIF) on cryosections of normal human ovarian tissue.

Non-human primate ovaries for the detection of AOA are not commercially available, while normal human ovarian tissue is available at the Institute of Pathology, Ljubljana, Slovenia, when ovaries are removed due to various pathological processes, particularly tumors, and are sent for pathohistological examination. Ovarian

tissue was incubated with patient serum diluted 1:10. The second incubation was with fluorescein isothiocyanate-labelled anti-human IgG antibody (Dakopatts, Copenhagen, Denmark). Positive reactions were semi-quantitatively evaluated (on a scale of 1–4+). Negative control omitting the patient's serum was regularly included (21). At the cellular level fresh PBMC were studied by flow cytometry (Becton Dickinson FACS, NJ, USA), and percentages of the following blood lymphocyte populations were determined: T cells (CD3⁺), helper T cells (CD4⁺), cytotoxic T cells (CD8⁺), natural killer cells (CD56+CD16+), regulatory T cells (CD25^{+high}) and B cells (CD19⁺) (22). Immunofluorescence labelling was performed by incubating PBMCs with monoclonal antibodies to CD3, CD4, CD8, CD25, CD56/16 and CD45. A differential blood count with a standard laboratory procedure was taken to obtain concentrations of individual lymphocyte subtypes.

Statistical analysis

Normal data distribution was tested with the Kolmogorov-Smirnov test. Where variables were normally distributed, we used the Pearson's chi-square test. If variables were not normally distributed, we used a nonparametric Mann-Whitney test. Statistical analysis was done using Statistical Package for the Social Sciences, version 18 (SPSS Inc., Chicago, IL, USA). The results were considered statistically significant at p<0.05.

Ethical considerations

The study protocol was approved by the National Ethics Committee, and all patients gave written informed consent.

Results

General clinical and biological parameters

The subjects were comparable with controls by age. One patient presenting a mosaic 45X0/46XX was excluded from the final analysis. Other patients had a 46XX karyotype. Anamnestic data of patients showed that four had had mumps during their childhood; they were thus excluded from the final analysis. Table 1 shows the clinical and endocrine characteristics of patients and of healthy women. All endocrine parameters in controls were within the normal range (Table 1).

Table 1: Hormone and ovarian peptide levels in patients and in healthy controls

Study group (n=20)	Control group (n=17)	
88.04 ± 47.93	6.25 ± 2.77	
37.85 ± 18.71	11.64 ± 31.1	
$180 \pm 200 \; (0.18 \pm 0.2 \; nmol/L)$	$210 \pm 130 \; (0.21 \pm 0.13 \; nmol/L)$	
11.14 ± 6.42	8.74 ± 5.51	
13.36 ± 10.66	32.63 ± 24.05	
0.36 ± 0.37	3.54 ± 1.58	
	88.04 ± 47.93 37.85 ± 18.71 $180 \pm 200 \ (0.18 \pm 0.2 \ \text{nmol/L})$ 11.14 ± 6.42 13.36 ± 10.66	

POI; Premature ovarian insufficiency, FSH; Follicle stimulating hormone, LH; Luteinizing hormone, E2; Estradiol, PRL; Prolactine and AMH; Anti-müllerian hormone.

Prevalence of autoimmune diseases and associated autoimmune abnormalities

We collected targeted history information on personal and familial autoimmune disorders (Table 2). Sixteen patients (80%) had an associated autoimmune disease in their personal and/or familial history. Four women (20%) with POI had first grade relatives with ovarian insufficiency before the age of 40. Thyroid disorders were the most common (15%) of the autoimmune diseases associated with POI in personal histories. Before

entering the study, three patients had been treated for autoimmune thyroid dysfunction (Hashimoto thyroiditis). In the study group, 55% of women had autoimmune disease in the family history; the most frequent autoimmune disorder was diabetes type I. The overall personal and familial incidence of autoimmune diseases was lower in the control group. In the study group, 50% of women had aTG. One healthy woman had evidence of autoimmune thyroid dysfunction, manifested by an elevated aTG serum level, and was excluded from the final analysis (Tables 2, 3).

Table 2: Personal and family history on autoimmune diseases in patients and healthy controls

	Study group (n=20)		Control group (n=17)	
	Family history	Personal history	Family history	Personal history
Diabetes type I	7		1	
Psoriasis	2	2	1	1
Rheumatism	2			
Thyroid disease		3	2	1
Vitiligo	1	1		
POI	4			
Atopic dermatitis		1		1

POI; Premature ovarian insufficiency.

Table 3: Associated autoimmune abnormalities (some participants showed more than one condition)

Associated autoimmune findings	Study group	Control group
Anti-thyroglobulin antibody (aTG)	10 (50%)	0
Positive ACTH test	5 (25%)	0
Anti-ovarian antibodies (AOA)	4 (20%)	0
Personal history of autoimmune disease	6 (30%)	2 (11.1%)
Family history of autoimmune disease	11 (55%)	4 (22.2%)
POI in family members	4 (20%)	0

POI; premature ovarian insufficiency.

Anti-ovarian antibodies

Analysis of AOA in serum was performed for all patients (Fig 1). AOA were detected in 4 (20%) patients, and in 3 patients, we found an associated autoimmune disease; Hashimoto thyroiditis. There was a clear positive immune fluorescent reaction of AOA in the serum of one woman who had mumps during her childhood, and she was excluded from the analysis. AOA were not detected in any of the controls (Table 3).

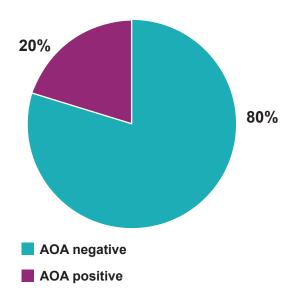


Fig 1: Prevalence of serum anti-ovarian antibodies (AOA) in patients.

Lymphocyte subtypes

Cellular autoimmune reaction occurring due to a changed T cell function was analysed as a potential cause of POI. Cell abnormalities were more frequent in women with POI than in healthy women (Table 4).

Peripheral T cell count is expressed as a percentage of various cell surface markers. In patients with POI, peripheral regulatory T lymphocytes (CD25^{+high}, p=0.015) were low and peripheral blood B cells (CD19⁺) were high (p=0.014); T lymphocyte parameters were normal in the control group.

Table 4: Prevalence (%) of analysed peripheral blood lymphocyte samples for various cell surface markers in patients and in controls

Various cell surface markers	Study group (%)	Control group (%)	P value
CD3 ⁺	73.25	74.53	0.578
CD19 ⁺	11.90	9.41	0.014
CD4 ⁺	47.45	45.82	0.450
CD8 ⁺	25.85	28.53	0.229
CD25 ^{+high}	1.60	2.85	0.015
CD56 ⁺ CD16 ⁺	14.20	16.23	0.366

Markers for peripheral blood lymphocytes: T lymphocytes (CD3⁺), helper T lymphocytes (CD4⁺), cytotoxic T lymphocytes, (CD8⁺), natural killer cells (CD56⁺CD16⁺), regulatory T lymphocytes (CD25^{+high}) and B cells (CD19⁺).

Discussion

To present POI as a possible autoimmune abnormality, we focused on three potentially interconnected factors: personal and familial history of autoimmune disorders, peripheral blood T-lymphocytes levels and presence of AOA.

We performed a targeted history of personal and familial autoimmune disorders and found associated autoimmune disorders in our patients. Thyroid disorders were common in personal histories and diabetes type 1 in familial histories, confirming the findings of previous studies (5, 6, 15). One of the reasons for suspecting an autoimmune etiology of POI is its frequent association with nearly all organ-specific autoimmune diseases (1, 3). Autoimmune diseases are significantly more frequent in young women than in men. This phenomenon may be explained by the effect of sex steroids on the components of the cellular immune system, which might contribute to the development and progression of autoimmune POI (23).

Hoek et al. (24) found that 60% of patients with secondary amenorrhea and Addison's disease have a detectable SCA serum titre. These antibodies are shown by 60-80% of patients with APGS type I. In 25% of our patients, the Synacthen test revealed an abnormal cortisol response. We interpreted this as the possibility that patients could be positive for SCA, but had a normal cortisol response. Patients with this disorder have an insidious onset; to confirm subclinical autoimmune adrenal insufficiency, measurement of adrenal antibodies might be a more effective screening method (25, 26).

Thyroid disorders and the presence of antithyroid antibodies are often mentioned in association with POI. Thyroid disorders are the most common of the autoimmune diseases associated with POI, found in 12-39% of women with POI (27-29). Thyroid disorders are often associated with endometriosis and polycystic ovary syndrome, two conditions often resulting in infertility (2). We found a greater involvement of aTG and aTPO in the study than in the control group, and the explanation being that ovarian failure may have been present in the latent period of thyroid disease. We found a strong correlation between autoimmune thyroid disease and autoimmune POI; 50% of patients tested positive for aTG. We concluded that not all patients with positive aTG necessarily have a clinically expressed thyroid disorder and the disease can have an insidious onset. Furthermore, greater involvement of other immune-mediated diseases, particularly auto immune disorders, such as allergy, psoriasis, atopic dermatitis and vitiligo in the study group suggests an autoimmune cause of POI, consistent also with the findings in the literature (9, 15, 30, 31).

Genetic predisposition is known to be one of the primary causes of autoimmunity and it is generally observed that patients with autoimmune diseases have several types of antibodies, as also confirmed in our study. We found AOA in 20% of patients: these results are consistent with other studies (9, 21, 32-35). The prevalence of AOA in women with POI, studied with IF on ovary tissue, varies greatly from 2 to 70%, which supports their role as a marker of an immune dysfunction process against ovaries. There could be several reasons for the differences among study results, including the diverse origin of tissue sections, which include human and non-human primate ovaries, as well as different study inclusion criteria. Many studies have shown that the presence of serum AOA does not correlate with the clinical manifestation of POI. Despite these antibodies being present, their pathogenetic role is highly questionable (36, 37). AOA may occur several years before the occurrence of clinical symptoms, as detected in 33-61% of women with unexplained infertility; a situation that may indicate early stages of autoimmune ovarian insufficiency (38, 39). For most autoimmune diseases, screening for specific antibodies is probably the best way of evaluating immunological involvement. Many ovarian structures are potential targets for autoimmune events with POI. In addition to AOA, SCA, aTG and aTPO, antibodies to several potential ovarian antigens have been proposed as markers of ovarian autoimmunity, which could potentially mediate autoimmune damage in POI. Betterle et al. (40) found antibodies against steroid genetic enzymes in some cases with anti-adrenal autoimmunity. Antibodies against gonadotropin receptors and gonadotropins have also been found in patients with POI, but they are still the subject of research. The general opinion is that more research is needed (41, 42). Although some studies have managed to identify autoantibodies against zona pellucida, corpus luteum and ovarian cells, these findings have no real correlation with the clinical picture (43-45). In our preliminary study, AOA were determined semi-quantitatively by the IIF method. IIF is a basic method and widely used for determining auto antibodies, while more specific tests enabling detection of AOA for specific antigen targets are not available in our laboratory.

Infertile women with AOA also have a decreased response to gonadotropin stimulation and reduced pregnancy rate after treatment (46, 47). In 2002, it was found that low-responders to gonadotropins with AOA are younger than low-responders without AOA (48). Detection of autoimmune processes that affect the ovarian response should thus be included in the diagnostic workup before any infertility treatment, particularly in women with low or no response to gonadotropins (34). Determination of AOA, as a specific test, is important in the diagnosis of diseases with an autoimmune etiology, but it should not be the only reliable diagnostic tool for optimal selection of patients who may benefit from immune modulatory therapy that could, at least temporarily, re-establish their ovarian function and fertility.

Abnormalities of cellular immunity of T lymphocytes, macrophages and dendritic cells play an important role in autoimmune events. Some of these abnormalities have been seen in women with POI, confirming the potential existence of an autoimmune mechanism of the disease. Mignot et al. (49) found that the absolute number and percentage of peripheral blood T lymphocytes, especially CD4⁺ T cells, are increased in patients with POI. Moreover, Miyake et al. (50) found that patients with POI have low levels of CD8+/CD57+ Tcells (cytotoxic T lymphocytes) and an increased ratio of CD4⁺ to CD8⁺ cells, which may reflect cell cytotoxic cell migration from blood to inflamed tissue. Multiple animal studies have suggested that the basis of POI is a cell-mediated autoimmune reaction caused by an alteration in T cell regulation (18). CD4⁺ T cells with constantly expressed α chain (CD25) receptor for IL-2 of were the first detected mediators of inhibition of autoimmune diseases in mice alteration of suppressor T cell subsets and T cell abnormalities are likely to play an important role in the pathogenesis of autoimmune diseases (51). Regulatory CD4+25+ T cells show a potent immunosuppressive function in vitro and in vivo, and contribute to immunologic self-tolerance by suppressing potentially auto-reactive CD4⁺ T cells. There is known to be a number of immunological

mechanisms associated with the failure of immune tolerance and the development of autoimmunity. Although studying regulatory T cells in human autoimmune diseases is difficult, and at times, findings have been contradictory, the data suggest that defects in CD4⁺ CD25⁺ regulatory T cells mediated suppression (52) are a major subset of immune cells responsible for peripheral immune self-tolerance.

In agreement with studies in patients with systemic lupus erythematosus, multiple sclerosis, rheumatoid arthritis and autoimmune vasculitis, we confirmed a reduced number of CD4+CD25+high T cells in the peripheral blood of our patients. High expressions of CD25 and CD4 surface markers have classically been used for identification of regulatory T cells. This may be problematic since CD25 is also expressed on antigen-responding activated non-regulatory T cells. The additional measurement of cellular expression of Foxp3 protein allowed a more specific analysis of Treg cells (CD4⁺CD25⁺Foxp3⁺ cells). However, Foxp3 is also transiently expressed in activated human effector T cells, thus complicating a correct Treg analysis. The large majority of Foxp3-expressing regulatory T cells express high levels of the interleukin-2 receptor alpha chain (CD25). Since there are no cell surface markers that are uniquely and specifically expressed on all Foxp3-expressing regulatory T cells, the measurement of CD4+CD25+high T cells is still in use in clinical studies of peripheral blood lymphocytes.

We interpreted the reduced number of CD4⁺CD25^{+high} T cells as a possible mechanism contributing to the formation of an autoimmune response in association with the presence of AOA and aTG. We also found an increased number of B cells in peripheral blood. A similar picture has been observed with other autoimmune endocrinopathies; therefore, we interpreted the elevated B cell count as activation of the humoral immune system, crucial for autoantibody production. Some authors, though, have tried estrogen substitution in women with POI without any effect on peripheral B cell count (53, 54).

The hormones inhibin B, FSH and AMH have been proposed as potential markers for determining the functional ovarian reserve (55, 56). In young ovulatory women, measurement of AMH at 3-year intervals has shown that the AMH serum

level decreases significantly over time, whereas other markers associated with ovarian aging, such as FSH, inhibin B and antral follicle count (AFC), do not change during this time period. Since it decreases at a time when concentrations of FSH and inhibin B are still normal, AMH has been proposed as the best indicator of ovarian reserve and as a marker of ovarian aging (55-59). In contrast, AMH is almost undetectable in women with POI (60), which our study also confirmed. In our group of patients, there was a small AFC or these structures were no longer seen on ultrasound, which agrees with the data in the literature (61).

Conclusion

Clinical and biological characteristics of women without known causes of disease suggest a possibility of autoimmune pathogenesis. In some patients, a combination of various autoimmune processes has been found. The presence of AOA and anti-thyroid antibodies, together with abnormalities of cellular immunity, potentially represent an autoimmune mechanism of POI. There is thus an increasing need to find a reliable and simple diagnostic procedure to determine the true prevalence of autoimmune ovarian disease. In women with POI, more attention should be paid to evaluation of associated autoimmune disorders.

Acknowledgements

We declare that we had no financial support from any pharmaceutical or other commercial company. The study was financed from research funds from the University Medical Centre Ljubljana, Slovenia, project number: 20100434. There is no conflict of interest in this study.

References

- Conway GS. Premature ovarian failure. Br Med Bull. 2000; 56(3): 643-649.
- Goswami D, Conway GS. Premature ovarian failure. Horm Res. 2007; 68(4): 196-202.
- Anasti JN. Premature ovarian failure: an update. Fertil Steril. 1998; 70(1): 1-15.
- Conway GS, Kaltsas G, Patel A, Davies MC, Jacobs HS. Characterisation of idiopathic premature ovarian failure. Fertil Steril. 1996; 65(2): 337-341.
- Husebye ES, Løvås K. Immunology of Addison's disease and premature ovarian failure. Endocrinol Metab Clin North Am. 2009; 38(2): 389-405.
- Poppe K, Velkeniers B. Female infertility and the thyroid. Best Pract Res Clin Endocrinol Metab. 2004; 18(2): 153-165.

- Edassery SL, Shatavi SV, Kunkel JP, Hauer C, Brucker C, Penumatsa K, et al. Autoantigens in ovarian autoimmunity associated with unexplained infertility and premature ovarian failure. Fertil Steril. 2010; 94(7): 2636-2641.
- Welt CK. Autoimmune oophoritis in the adolescent. Ann NY Acad Sci. 2008; 1135: 118-122.
- Ashrafi M, Fallahian M, Eshrati B, Salman Yazdi R. The presence of anti thyroid and anti ovarian auto-antibodies in familial POF. Int J Fertil Steril. 2008; 1(4): 171-174.
- Irvine WJ, Barnes EW. Addison's disease, ovarian failure and hypoparathyroidism. Clin Endocrinol Metab.1975; 4(2): 379-434.
- Turkington RW, Lebovitz HE. Extra-adrenal endocrine deficiencies in Addison's disease. Am J Med. 1967; 43(4): 499-507
- Muir A, Schatz DA, MacLaren NK. Autoimmune polyglandular syndrome. In: de Groot LS, editor. Endocrinology. 3rd ed. Philadelphia: WB Saunders; 1995; 3013-3024.
- Blizzard RM, Chee D, Davies W. The incidence of adrenal and other antibodies in sera of patients with idiopathic adrenal insufficiency (Addison's disease). Clin Exp Immunol. 1967; 2(1): 19-30.
- Irvine WJ, Chan MM, Scarth L, Kolb FO, Hartog M, Bayliss RI, et al. Immunological aspects of premature ovarian failure associated with idiopathic Addison's disease. Lancet. 1968; 2(7574): 883-887.
- Forges T, Monnier-Barbarino P, Faure GC, Bene MC. Autoimmunity and antigenic targets in ovarian pathology. Hum Reprod Update. 2004; 10(2): 163-175.
- 16. Morita Y, Tilly JL. Oocyte apoptosis: like sand through an hourglass. Dev Biol. 1999; 213(1): 1-17.
- 17. Zuckerman S. The number of oocytes in the mature ovary. Recent Prog Horm Res. 1951; 6(1): 63-108.
- Byskof AG, Faddy MJ, Lemmen JG, Andersen CY. Eggs forever?. Differentiation. 2005; 73(9-10): 438-446
- Liu Y, Wu C, Lyu Q, Yang D, Albertini DF, Keefe DL, et al. Germline stem cells and neo-oogenesis in the adult human ovary. Dev Biol. 2007; 306(1): 112-120.
- Bukovsky A, Virant-Klun I, Svetlikova M, Willson I. Ovarian germ cells. Methods Enyzmol. 2006; 419: 208-258.
- 21. Damewood MD, Zacur HA, Hoffman GJ, Rock JA. Circulating antiovarian antibodies in premature ovarian failure. Obstet Gynecol. 1986; 68(6): 850-854.
- Groselj-Grenc M, Ihan A, Derganc M. Neutrophil and monocyte CD64 and CD163 expression in critically ill neonates and children with sepsis: comparison of fluorescence intensities and calculated indexes. Mediators Inflamm. 2008; 2008: 202646.
- Lahita RG. Predisposing factors to autoimmune disease. Int J Fertil Womens Med. 1997; 42(2): 115-119.
- Hoek A, Schoemaker J, Drexhage HA. Premature ovarian failure and ovarian autoimmunity. Endocr Rev. 1997; 18(1): 107-134.
- Bakalov VK, Vanderhoof VH, Bondy CA, Nelson LM. Adrenal autoantibodies detected asymptomatic autoimmune adrenal insufficiency in young women with spontaneous premature ovarian failure. Hum Reprod. 2002; 17(8): 2096-2100.
- Brozzetti A, Marzotti S, La Torre D, Bacosi ML, Morelli S, Bini V, et al. Autoantibody responses in autoim-

- mune ovarian insufficiency and in Addison's disease are IgG1 dominated and suggest a predominant, but not exclusive, Th1 type of response. Eur J Endocrinol. 2010; 163(2): 309-317.
- Dragojevic-Dikic S, Marisavljevic D, Mitrovic A, Dikic S, Jovanovic T, Jankovic-Raznatovic S. An immunological insight into premature ovarian failure (POF). Autoimmun Rev. 2010; 9(11): 771-774.
- Betterle C, Rossi A, Dalla Pria S, Artifoni A, Pedini B, Gavasso S, et al. Premature ovarian failure: autoimmunity and natural history. Clin Endocrinol. 1993; 39(1): 35-43.
- Goswami R, Marwaha RK, Goswami D, Gupta N, Ray D, Tomar N, et al. Prevalence of thyroid autoimmunity in sporadic idiopathic hypoparathyroidism in comparison to type 1 diabetes and premature ovarian failure. J Clin Endocrinol Metab. 2006; 91(11): 4256-4259.
- 30. Irvine WJ. Premature menopause in autoimmune disease. Lancet. 1969; 1(7588): 264.
- Vallotton MB, Forbes AP. Antibodies to cytoplasm of ova. Lancet. 1966; 2(7457): 264-265.
- Gleicher N, Weghofer A, Oktay K, Barad D. Do etiologies of premature ovarian aging (POA) mimic those of premature ovarian failure (POF)?. Hum Reprod. 2009; 24(10): 2395-2400.
- Goswami D, Conway GS. Premature ovarian failure. Hum Reprod Update. 2005; 11(4): 391-410.
- Luborsky J. Ovarian autoimmune disease and ovarian autoantibodies. J Womens Health Gend Based Med. 2002; 11(7): 585-599.
- Yan G, Schoenfeld D, Penney C, Hurxthal K, Taylor A, Faustman D. Identification of premature ovarian failure patients with underlying autoimmunity. J Womens Health Gend Based Med. 2000; 9(3): 275-287.
- Monnier-Barbarino P, Forges T, Faure GC, Béné MC. Gonadal antibodies interfering with female reproduction. Best Pract Res Clin Endocrinol Metab. 2005; 19(1): 135-148.
- Coulam CB, Ryan RJ. Prevalence of circulating antibodies directed towards ovaries among women with premature ovarian failure. Am J Reprod Immunol Microbiol. 1985; 9(1): 23-24.
- Gleicher N, Weghofer A, Barad D. Female infertility due to abnormal autoimmunity: frequently overlooked and greatly underappreciated. Part II. Expert Rev Obstet Gynecol. 2007; 2(4): 465-475.
- Gleicher N, Weghofer A, Barad D. Female infertility due to abnormal autoimmunity: frequently overlooked and greatly underappreciated. Part I. Expert Rev Obstet Gynecol. 2007; 2(4): 453-464.
- Betterle C, Rossi A, Dalla Pria S, Artifoni A, Pedini B, Gavasso S, et al. Premature ovarian failure: autoimmunity and natural history. Clin Endocrinol. 1993; 39(1): 35-43.
- Wheatcroft NJ, Toogood AA, Li TC, Cooke ID, Weetman AP. Detection of antibodies to ovarian antigens in women with premature ovarian failure. Clin Exp Immunol. 1994; 96(1): 122-128.
- Luborsky JL, Visintin I, Boyers S, Asari T, Caldwell B, DeCherney A. Ovarian antibodies detected by immobilized antigen immunoassay in patients with premature ovarian failure. J Clin Endocrinol Metab. 1990; 70(1): 69-75.
- 43. Moncayo H, Moncayo R, Benz R, Wolf A, Lauritzen

- C. Ovarian failure and autoimmunity: detection of autoantibodies directed against both the unoccupied luteinizing hormone/human chorionic gonadotropin receptor and the hormone-receptor complex of bovine corpus luteum. J Clin Invest. 1989; 84(6): 1857-1865.
- 44. Sotsion F, Bottazzo GF, Doniach D. Immunoofluorescence studies on antibodies to steroid-producing cells, and to germline cells in endocrine disease and infertility. Clin Exp Immunol. 1980; 39(1): 97-111.
- Horejsi J, Martinek J, Novakova D, Madar J, Brandejska M. Autoimmune antiovarian antibodies and their impact on the success of an IVF/ET program. Ann N Y Acad Sci. 2000; 900: 351-356.
- Meyer WR, Lavy G, DeCherney AH, Visintin I, Economy K, Luborsky JL. Evidence of gonadal and gonadotropin antibodies in woman with suboptimal ovarian response to exogenous gonadotropin. Obstet Gynecol. 1990; 75(5): 795-799.
- 47. Luborsky J, Pong R. Pregnancy outcome and ovarian antibodies in infertility patients undergoing controlled ovarian hyperstimulation. Am J Reprod Immunol. 2000; 44(5): 261-265.
- Luborsky JL, Thiruppathi P, Rivany B, Roussev R, Coulam C, Radwanska E. Evidence for different etiologies of low estradiol response to FSH: age-related accelerated luteinization of follicles of presence of ovarian autoantibodies. Hum Reprod. 2002; 17(10): 2641-2649.
- Mignot MH, Drexhage HA, Kleingeld M, Van de Plassche-Boers EM, Rao BR, Schoemaker J. Premature ovarian failure. II: Considerations of cellular immunity defects. Eur J Obstet Gynecol Reprod Biol. 1989; 30(1): 67-72.
- Miyake T, Sato Y, Takeuchi S. Implications of circulating autoantibodies and peripheral blood lymphocyte subset for the genesis of premature ovarian failure. J Reprod Immunol. 1987; 12(3): 163-171.
- Sakaguchi S, Sakaguchi N, Asano M, Itoh M, Toda M. Immunologic self-tolerance maintained by activated T cells expressing IL-2 receptor alpha-chains (CD25). Breakdown of a single mechanism of self-tolerance causes various autoimmune diseases. J Immunol. 1995; 155(3): 1151-1164.
- 52. Long SA, Buckner JH. CD4+FOXP3+ T regulatory cells in human autoimmunity: more than a numbers game. J Immunol. 2011; 187(5): 2061-2066.
- Chernysmov VP, Radysh TV, Gura IV, Tatarchuk TP, Khominskaya ZB. Immune disorders in women with premature ovarian failure in initial period. Am J Reprod Immunol. 2001; 46(3): 220-225.
- 54. Ho PC, Tang GW, Lawton JW. Lymphocyte subsets and serum immunoglobulins in patients with premature ovarian failure before and after estrogen replacement. Hum Reprod. 1993; 8(5): 714-716.
- 55. Van Houten EL, Themmen AP, Visser JA. Anti-Müllerian hormone (AMH): regulator and marker of ovarian function. Ann Endocrinol (Paris). 2010; 71(3): 191-197.
- Baird DD, Steiner AZ. Anti-Müllerian hormone: a potential new tool in epidemiologic studies of female fecundability. Am J Epidemiol. 2012; 175(4): 245-249.
- 57. Van Rooij IA, Broekmans FJ, Scheffer GJ, Looman CW, Habbema JD, de Jong FH, et al. Serum anti müllerian hormone levels best reflect the reproductive decline with age in normal women with proven

Košir Pogačnik et al.

- fertility: a longitudinal study. Fertil Steril. 2005; 83(4): 979-987.
- Domingues TS, Rocha AM, Serafini PC. Tests for ovarian reserve: reliability and utility. Curr Opin Obstet Gynecol. 2010; 22(4): 271-276.
- Meden-Vrtovec H, Osredkar J. Anti-Müllerian hormone-a predictor of ovarian reserve. Zdrav Vestn. 2010; 79(6): 507-511.
- 60. Nelson SM, Anderson RA, Broekmans FJ, Raine-Fen-
- ning N, Fleming R, La Marca A. Anti-Müllerian hormone: clairvoyance or crystal clear?. Hum Reprod. 2012; 27(3): 631-636.
- 61. Knauff EA, Eijkemans MJ, Lambalk CB, ten Kate-Booij MJ, Hoek A, Beerendonk CC, et al. Anti-Müllerian hormone, inhibin B, and antral follicle count in young women with ovarian failure. J Clin Endocrinol Metab. 2009; 94(3): 786-792.